

Identification of Unique patterns, Mutation in Genetic finger printing using neural network approach

Mr.T.Saravanan¹,Head & Assistant Professor, Dr.B.Mukunthan², Associate Professor, PG & Research Department of Computer Science, Jairams Arts & Science College, Karur-639003.

Abstract – In genetic engineering, the advent of human genome project immensely increased the pressure for molecular computations and sequencing technologies dealing with data beyond the current abilities that are to be sequenced and interpreted. The automation of DNA feature extraction process achieved by applying neural network technique which has the advantage over conventional programming, in their ability to solve problem that do not have an algorithmic solution or the available solutions is too complex to be found is discussed in this paper, the above work also reduces the complication in precisely analyzing, interpreting of human DNA. The identification of exact location of occurrence of mutation in the DNA chain caused by radiation, viruses, transposons, mutagenic chemicals, as well as errors occurring during meiosis or DNA replication can be easily and exactly detected by subjecting the sample to gene sequencing process and analyzed using the above technique. In this novel approach the perfect blend made of bioinformatics and neural networks technique results in efficient DNA pattern analysis algorithm with 100% prediction accuracy, computed by number of correct identification of the target for a set of given inputs.

Key words - Neural-Fuzzy Resonance Mapping, Competitive learning, NFPR-processor, Input Generator, Preprocessor, Separator, Discriminator and Comparator, DNA profiling, DNA sequence Format, Mutation.

1. INTRODUCTION

Knowledge of DNA sequences has become indispensable for basic biological research. DNA sequencing is applied in various fields such as diagnostic, biotechnology, forensic biology and biological systematic. The DNA sequences of thousands of organisms have been decoded and stored in databases. The sequence information is analysed to determine genes that



encode polypeptides, RNA genes, regulatory sequences, structural motifs, and repetitive sequences. A comparison of genes within a species or between different species can show similarities between protein functions, or relations between species. With the growing amount of data, it became impractical to analyse DNA sequences manually.

Neural networks learn by examples so that it can be trained with known examples of a problem to gain knowledge about it so the neural network can be effective to solve unknown or untrained instances of the problem if is aptly trained. A pattern is essentially an arrangement or an ordering, in which some organization of underlying structure can be said to exist i.e. a pattern can be referred to as a quantitative or structural description of an object or some item of interest. A set of patterns that share some common properties can be regarded as pattern class in our case the unique repeated nucleotide sequence from the given Human DNA sample. The concept of applying Artificial Neural Systems (ANS) or Artificial Neural Networks (ANN) or simply Neural Networks in the field of DNA profiling is discussed in this paper.

2. ARTIFICAL NEURAL NETWORK TECHNIQUES

Neural Networks [3] can process information in parallel, at high speed, and in a distributed manner. Neural networks which are simplified models of the biological neuron system, is a massively parallel distributed processing system made up of highly interconnected neural computing elements that have the ability to learn and thereby acquire knowledge and make it available for use. Neural Network architectures have been classified into various types based on their learning mechanisms and other features. Some classes of Neural Network refer to this learning process as training and the ability to solve a problem using the knowledge acquired as inference.

Neural Networks exhibit mapping capabilities, i.e., they can map input patterns to their associated output patterns. Neural Networks architectures can be trained with known examples of a problem before they are tested for their inference. They can, therefore, identify new objects previously untrained. Neural Networks possess the capability to generalize i.e. they can



predict new outcomes from past trends. Neural Networks are robust systems and are fault tolerant. They can therefore, recall full patterns from incomplete, partial or noisy patterns.

In Competitive Learning method those neurons which respond strongly to input stimuli have their weights updated, when an input pattern is presented, all neurons in the layer compete and the winning neuron undergoes weight adjustment. Hence it is a "Winner-takes-all" strategy.

Adaptive resonance theory employs a new principle of self organization based on competitive learning .Adaptive resonance theory nets are designed to be both stable and plastic. Neural networks suitable particularly for pattern classification problems in realistic environment is Neural- Fuzzy resonance mapping , it is a vast simplification of fuzzy resonance mapping which possess reduced computational overhead and architectural redundancy when compared to fuzzy resonance mapping.

3. DNA PROFILING AND SEQUENCING

DNA profiling also called DNA testing, DNA typing, or genetic fingerprinting, is a technique employed by forensic scientists to assist in the identification of individuals on the basis of their respective DNA profiles. DNA profiles, are encrypted sets of numbers that reflect a person's DNA makeup, which can also be used as the person's identifier. DNA sequencing theory addresses physical processes related to sequencing DNA .The term DNA sequencing [19] refers to sequencing methods for determining the order of the nucleotide bases—adenine, guanine, cytosine, thymine and uracil(rare case) in a molecule of DNA.

Single nucleotide poly-orphisms are a DNA sequence variation occurring when a single nucleotide A, T, C, or G in the genome (or other shared sequence) differs between members of a species (or between paired chromosomes in an individual). The genome [21] is the entirety of an organism's hereditary information which is encoded either in DNA or, for many types of virus, in RNA. For example, two sequenced DNA fragments from different individuals, AAGCCTA to AAGCTTA, contain a difference in a single nucleotide. Various DNA Sequence Formats available are:1) Plain sequence format 2) EMBL format 3) GCG format 4) GCG-RSF (rich sequence format 5) Gen Bank format 6) IG format 7) FASTA format. A





sequence file in FASTA format of a given sample is used as an input to that is to be interpreted and analysed.

4. NEURAL-FUZZY PATTERN RECOGNITION PROCESSOR

4.1 Learning Input Generator

The input generator is used for input normalization and it represents the presence of particular feature in the input patterns and its absence. Various conditions for generating normalized learning input are shown below.

Learning Inputs

 $LIN_{i,n} = I_1, I_2, ..., I_p$

(1)

Where	$0.1 \le i$	$\leq 0.5, \ 0.1 \leq r$	<mark>1 ≤ 0.5</mark>
and	p = 4		

TABLE 1

CONDITIONS FOR LEARNING INPUT NORMALIZATION

	Condition	Learning Input	Category
Case 1	i ≠ n or i=n=0.1 and n<=0.5	LIN _{i, n} = i, n, 1- i, 1-n e.g. LIN _{0.1, 0.1} = 0.1, 0.1, (1- 0.1), (1-0.1) LIN _{0.1, 0.1} = 0.1, 0.1, 0.9, 0.9 LIN _{0.2, 0.5} = 0.2, 0.5, (1- 0.2), (1-0.5) LIN _{0.2, 0.5} = 0.2, 0.5, 0.8, 0.5	Category=L(logical)
Case 2	i = n and 0.1> i, n <0.5	LIN _{i, n} = i, 1-i, 1-n, n e.g. LIN _{0.2, 0.2} = 0.2, (1-0.2), (1-0.2), 0.2 LIN _{0.2, 0.2} = 0.2, 0.8, 0.8,	Category=ILL(illogic al)



		0.2 LIN $_{0.3, 0.3}=$ 0.3, (1-0.3), (1-0.3), 0.3 LIN $_{0.3, 0.3}=$ 0.3, 0.7, 0.7, 0.3	
Case 3	i≠n and n>0.5	LIN _{i, n} = i, n, 1- i, 1-n e.g. LIN _{0.5, 0.6} = 0.5, 0.6, (1- 0.5), (1- 0.4) LIN _{0.5, 0.6} = 0.5, 0.6, 0.5, 0.4	Category=ILL (illogical)

4.2 Activatio<mark>n Func</mark>tion Generator

(2)

When coded input patterns from input generator are presented to NFPR-Processor all output nodes become active to varying degrees. The output activation denoted by ACFj referred to as the activation function for the jth output node. Where LIN is the learning input and LIW_j is the corresponding learning input weights.

$$ACFj = \frac{\left| LIN \land LI Wj \right|}{\alpha + \left| LI Wj \right|}$$

Here α is kept as a small value close to 0 it's about 0.0000001. The node which registers the highest activation function is deemed Winner node i.e.

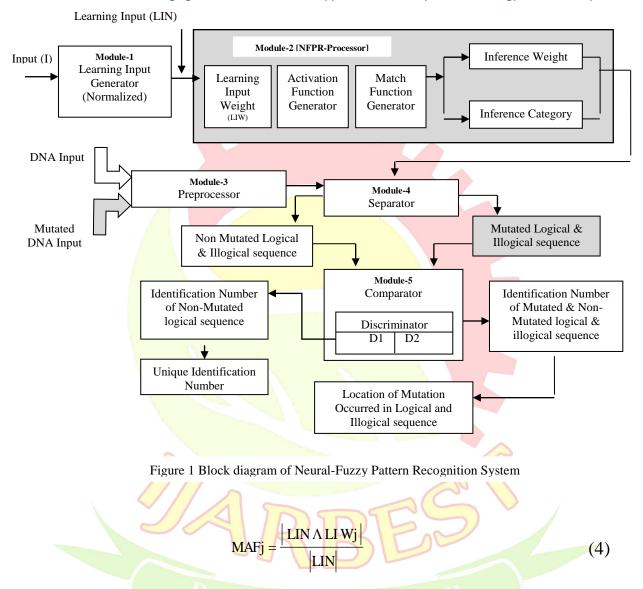
Winner node =
$$max(ACFj)$$
 (3)

In the event of more than one node emerging as the winner owing to the same activation function value some mechanism such as choosing a node with the smallest index may be devised to break the tie.

4.3 Match Function Generator

The match function which helps to determine whether the network must adjust its learning parameters is given by





The match function in association with the vigilance parameter decides on whether a particular output node is good enough to encode a given input pattern or whether a new output node should be opened to encode the same. The network is said to be in a state of resonance, if the match function value exceeds vigilance parameter. However, for a node to exhibit resonance, it is essential that it not only encodes the given input pattern but should also represent the same category as that of the input pattern.



The network is said to be in state of mismatch reset if the vigilance parameter exceeds match function, Such a state only means that the particular output node is not fit enough to learn the given input pattern and thereby cannot update its weights even though the category of the output node may be the same as that of the input pattern. This is so, since the output node has fallen short of the expected encoding granularity indicated by the vigilance parameter.

If match function is greater than vigilance parameter and category of input pattern is not same with the learning input, the vigilance parameter is updated and is given by

 $\rho = MAF + \delta$ ($\delta = 0.001$)

(5)

		TABLE 2			
GENERA	ATING WEIGHT	S FOR IN	FERENCE,	CATE	EGORY FOR
	INFERENCE	FROM LE	ARNING I	NPUT.	S

Nucleotid e Pair	A,A	A,U	T,A	T,T	T, U	G,A	G, G	G,U	C,A	C,C	C,U	U,A	U,U
Category	L	L	L	ILL	L	L	ILL	E	L	ILL	L	L	ILL
Fuzzy Equivalent	0.1, 0.1*	0.1, 0.5*	0.2, 0.1*	0.2, 0.8* *	0.2 ,0. 5*	0.3, 0.1*	0.3, 0.7* *	0.3, 0.5*	0.4, 0.1*	0.4, 0.6* *	0.4, 0.5*	0.5, 0.1*	0.5,0.6 ***
Compleme nt of Learning Input	0.9, 0.9	0.9, 0.5	0.8, 0.9	0.8, 0.2	0.8 ,0. 5	0.7, 0.9	0.7, 0.3	0.7, 0.5	0.6, 0.9	0.6, 0.4	0.6, 0.5	0.5, 0.9	0.5,0.4
Augmente d Input / Learning Input(LI)	0.1, 0.1, 0.9, 0.9	0.1, 0.5, 0.9, 0.5	0.2, 0.1, 0.8, 0.9	0.2, 0.8, 0.8, 0.2	0.2 ,0. 5, 0.8 ,0. 5	0.3, 0.1, 0.7, 0.9	0.3, 0.7, 0.7, 0.3	0.3, 0.5, 0.7, 0.5	0.4, 0.1, 0.6, 0.9	0.4, 0.6, 0.6, 0.4	0.4, 0.5, 0.6, 0.5	0.5, 0.1, 0.5, 0.9	0.5,0.6, 0.5,0.4



International Journal of Advanced Research in Biology Ecology Science and Technology (IJARBEST) Vol. 2, Special Issue I, January 2016 in association with KARUR VELALAR COLLEGE OF ARTS AND SCIENCE FOR WOMEN, KARUR

Conference on Emerging Trends and Functional Applications of Computer Technology - 12th January 2016

δ)	0.00	0.00	0.00	0.00	0.0	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.001
		1	1	1	1	01	1	1	1	1	1 0.5,	1	1	
ρ)	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.3, 0.60 0+ δ	0.60 1	0.60 1	0.601
β	<u> </u>	1	1	1	1	1	1	1	1	1	1	1	1	1
Acti	AC F(1)	0.99 99	0.79 99	0.93 75	0.79 99	0.9 99 9	0.93 33	0.87 51	0.99 99	0.92 85	0.92 30	0.99 99	0.92 30	0.8461
vatio n Func	AC F(2)	2	7	~	4	0.8 49 9	0.59 99	0.89 99	0.88 88	0.61 11	0.88 88	0.93 75	0.62 49	0.9999
tion	AC F(3)	2	1	~	2	۲	~	~	~	~	2	~	~	0.7499
Higl Activa Func	nest ation	AC F(1)	AC F(1)	AC F(1)	AC F(2)	AC F(1)	AC F(1)	AC F(2)	AC F(1)	AC F(1)	AC F(2)	AC F(1)	AC F(1) , AC F(2)	ACF(2)
Mat	MA F(1)	1.00 00	0.80 00	0.75 00	0.60 00	0.7 50 0	0.70 00	0.60 00	0.70 00	0.65 00	0.60 00	0.65 00	0.60 00	0.4500
ch Func tion	MA F(2)	2	3	~~~~		0.8 50 0	0.60	0.90 00	0.80	0.55 00	0.80 00	0.75 00	0.50 00	0.7500
	MA F(3)	2	ł	~	lese	w	i at	its I	Best	11	~	~	۲	0.700
Cate	•	Mat ch	Mat ch	Mat ch		Ma tch	Mat ch		Mat ch	Mat ch	Mis mat ch	Mat ch	Mat ch	
Mat Mism					Mat ch			Mat ch			Mat ch			Match
Lear ning	LI W(=LI	Up dat	Up dat	Not Up	Up dat	Up dat	Not Up	Up dat	Up dat	Not Up	Up dat	Not Up	Not Updat

25



International Journal of Advanced Research in Biology Ecology Science and Technology (IJARBEST) Vol. 2, Special Issue I, January 2016 in association with KARUR VELALAR COLLEGE OF ARTS AND SCIENCE FOR WOMEN, KARUR

Conference on Emerging Trends and Functional Applications of Computer Technology - 12th January 2016

Inpu	1)		ed	ed	dat	ed	ed	dat	ed	ed	dat	ed	dat	ed
t					ed			ed			ed		ed	
Wei													(<p)< td=""><td></td></p)<>	
ghts						No	Not		Nat	Not		Nat	Not	
Upd	LI					t	Not	Up	Not	Not	Up	Not	Up	
ated/	W (~	~	~	=LI	Up	Up	dat	Up	Up	dat	Up	dat	Updat
Not	2)					dat	dat	ed	dat	dat	ed	dat	ed	ed
Upd						ed	ed		ed	ed		ed	(<p)< td=""><td></td></p)<>	
ated/	LI					and the second second	ander Constantion							Not
Add	W (~	~	~	~	~	~	~	~	~	~	~	Ad	Updat
ed	3)												ded	ed
			j.			0.1								WFI(1
	LI	0.1,	0. <mark>1</mark> ,	0.1,	0.1,	,0.	0.1,	0.1,	0.1,	0.1,	0.1,	0.1,	0.1,)=0.1,0
	W (0.1,	0. <mark>1</mark> ,	0.1,	0.1,	1,	0.1,	0.1,	0.1,	0.1,	0.1,	0.1,	0.1,	.1,
	1)	0.9,	0.9,	0.8,	0.8,	0.8	0.7,	0.7,	0.7,	0.6,	0.6,	0.6,	0.6,	0.6,0.5
-	Ĺ	0.9	0.5	0.5	0.5	,0.	0.5	0.5	0.5	0.5	0.5	0.5	0.5	CFI (1)
Lear						5								=L
ning						0.2								WFI(2
Inpu	LI				0.2,	,0.	0.2,	0.2,	0.2,	0.2,	0.2,	0.2,	0.2,)=0.2,0
t .	W (0.8,	8,	0.8,	0.7,	0.7,	0.7,	0.6,	0.6,	0.6,	.5,
Wei	2)	~	~	S	0.8,	0.8	0.8,	0.7,	0.7,	0.7,	0.6,	0.6,	0.6,	0.5,0.2
ghts	ILL			$(/\gamma)$	0.2	,0.	0.2	0.2	0.2	0.2	0.2	0.2	0.2	CFI(2)
& Cata					1	2			20	5				=ILL
Cate			C	20	7	11-	\mathcal{Y}	221	5	\mathcal{I}				WFI(3
gory	LI							(C	2				0.5,)=0.5,0
	W (0.1,	.1,
	3)	~	~	~	200	~	~	~	~ t	Ĩ	~	~	0.5,	0.5,0.9
	L					arch) at	its I	sesu				0.9	CFI(2)
	A-ADENINE, T-THYMINE, G-GUANINE, C-CYTOSINE,U-													
	URACIL,ACF=ACTIVATION FUNCTION, MAF=MATCH FUNCTION, CFI= CATEGORY FOR INFERENCE, WFI= WEIGHT FOR INFERENCE , L=LOGICAL,													
					,							,		,
ILL=	ILL=ILLOGICAL ρ=VIGILANCE PARAMETER *- CASE1,**-CASE2,***CASE3,=LI-													
]	EQUA	L TO	LEAF	RNIN	G INP	°UT,<)=LES	S TH	AN RI	HO		



DNA INPUTS

The weight updating equation of an output node *j* when it proceeds to learn the given input pattern *I* is given by

WFI j^{new} = β (LIN \land WFIj^{old}) + (1 - β)WFIj^{old}

(6)

where $0 \le \beta \le 1$ $(\beta = 1)$

Once the network has been trained, the inference of patterns, logical or illogical i.e. the categories to which the patterns belong may be easily computed. This is accomplished by passing the input pattern into the preprocessor and then to the input layer. All the output nodes compute the activation functions with respect to the input. The winner, node with the highest activation function, is chosen. The category to which output node belongs is the one to which given input pattern is classified by the network.

PPO 🔨 WFIj

WFI

CIF_j =

(7) If CIF (1) or CIF (3) is greater than CIF (2) the inferred category is logical else if CIF (2) is greater than CIF (1) and CIF (3) then inferred category is illogical. For the DNA inputs of fast a format whose category is logical the corresponding seven consecutive nucleotide base in the DNA sample is chosen as single logical sequence and DNA inputs whose category is illogical, two consecutive nucleotide base is considered as an illogical sequence with base pair thirty two.

TABLE 3

GENERATING WEIGHTS FOR INFERENCE, CATEGORY INFERENCE FUNCTION FROM LEARNING INPUTS



International Journal of Advanced Research in Biology Ecology Science and Technology (IJARBEST) Vol. 2, Special Issue I, January 2016 in association with KARUR VELALAR COLLEGE OF ARTS AND SCIENCE FOR WOMEN, KARUR

Conference on Emerging Trends and Functional Applications of Computer Technology - 12th January 2016

		0.1,0	0.2,0	0.4,0	0.4,0	0.4,0	0.4,0	0.2,0	0.1,0	0.2,0	0.2,0	0.2,0	0.2,0
		.1	.3	.4	.4	.4	.2	.4	.1	.2	.2	.2	.1
		A,A	T,G	C,C	C,C	C,C	C,T	T,C	A,A	T,T	T,T	T,T	T,A
		0.1,0	0.2,0	0.4,0	0.4,0	0.4,0	0.4,0	0.2,0	0.1,0	0.2,0	0.2,0	0.2,0	0.2,0
PP	20	.1,	.3,	.6,	.6,	.6,	.2,	.4,	.1,	.8,	.8,	.8,	.1,
11	U	0.9,0	0.8,0	0.6,0	0.6,0	0.6,0	0.6,0	0.8,0	0.9,0	0.8,0	0.8,0	0.8,0	0.8,0
		.9	.7	.4	.4	.4	.8	.6	.9	.2	.2	.2	.9
		0.1,0	0.1,0	0.1,0	0.1,0	0.1,0	0.1,0	0.1,0	0.1,0	0.1,0	0.1,0	0.1,0	0.1,0
	WFI(.1,	.1,	.1,	.1,	.1,	.1,	.1,	.1,	.1,	.1,	.1,	.1,
	1)	0.6,0	0.6,0	0.6,0	0.6,0	0.6,0	0.6,0	0.6,0	0.6,0	0.6,0	0.6,0	0.6,0	0.6,0
		.5	.5	.5	.5	.5	.5	.5	.5	.5	.5	.5	.5
-		0.2,0	0.2,0	0.2,0	0.2,0	0.2,0	0.2,0	0.2,0	0.2,0	0.2,0	0.2,0	0.2,0	0.2,0
WFI	WFI(.5,	.5,	.5,	.5,	.5,	.5,	.5,	.5,	.5,	.5,	.5,	.5,
	2)	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0
		.2	.2	7.2	.2	.2	.2	.2	.2	.2	.2	.2	.2
		0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0
	WFI(.1,	.1,	.1,	.1,	.1,	$\mathbf{)}1$	<u>,</u> ,	.1,	.1,	.1,	.1,	.1,
	3)	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0	0.5,0
		.9	.9	.9	°a.9ci	1 .9t	it <u>s</u> 9 b	.9	.9	.9	.9	.9	.9
-		1.00	1.00	0.92	0.92	0.92	1.00	1.00	1.00	0.76	0.76	0.76	1.00
	CIF(1)	00	00	30	30	30	00	00	00	92	92	92	00
CIF	CIF(2	0.64	0.85	1.00	1.00	1.00	0.78	0.92	0.64	1.00	1.00	1.00	0.50
)	28	71	00	00	00	51	85	28	00	00	00	00
	CIF(3	0.80	0.75	0.70	0.70	0.70	0.90	0.70	0.80	0.85	0.85	0.85	0.85



International Journal of Advanced Research in Biology Ecology Science and Technology (IJARBEST) Vol. 2, Special Issue I, January 2016 in association with KARUR VELALAR COLLEGE OF ARTS AND SCIENCE FOR WOMEN, KARUR

Conference on Emerging Trends and Functional Applications of Computer Technology - 12th January 2016

)	00	00	00	00	00	00	00	00	00	00	00	00
	FIC	CIF(CIF	CIF	CIF	CIF	CIF	CIF	CIF	CIF	CIF	CIF	CIF(
G	πC	1)	(1)	(2)	(2)	(2)	(1)	(1)	(1)	(2)	(2)	(2)	1)
	LOGI CAL	L	L	5			L		L				L
IC	ILLO		1						5				
	GICA			ILL	ILL	ILL				ILL	ILL	ILL	
	L												
		0.1,0	0.2,0				0.4,0	0.2,0	0.1,0				0.2,0
		.1,0.	.3,0.				.2,0.	.4,0.	.1,0.				.1,0.
S	OP	2,0.3	<mark>2,0.3</mark>	0.4,0	0.4,0	0.4,0	4,0.1	<mark>2,0.4</mark>	2,0.3	0.2,0	0.2,0	0.2,0	4,0.1
5	Or	,0.2,	, <mark>0.2</mark> ,	.4	.4	.4	,0.1,	,0.2,	,0.2,	.2	.2	.2	,0.4,
		0.3,0	0. 3 ,				0.1,	0.4,	0.3,				0.2,
		.2	0.1				0.1	0.1	0.2	\sim			0.4
A=	ADENIN	NE,T=7		1 1		and the second s					EPROC	CESSO	R
	CIF=CA	TECC	11 - 1			EIGH		Norman State	and the second se	,	NFFD	BED	
		ALEGU				RY,IC			24			πĽD	
	CATEG	ORY,I	L=LOC	JICAL	,ILL=]	ILLOC	HCAL	,SOP=	SEPAI	RATO	R OUT	PUT	

Logical sequence (LS): LSp, s, k = Lseq p, s, 1, Lseq p, s, 2,...,Lseq p, s, k (8) Where p, s = 1 to ∞

and k = 1 to 7

The separator outputs which are logical in their category are fed to the discriminator (D1) where identification number is computed using the equation below to generate unique identification number



International Journal of Advanced Research in Biology Ecology Science and Technology (IJARBEST) Vol. 2, Special Issue I, January 2016 in association with KARUR VELALAR COLLEGE OF ARTS AND SCIENCE FOR WOMEN, KARUR Conference on Emerging Trends and Functional Applications of Computer Technology - 12th January 2016

$$D1_{p,s} = \sum_{k=1}^{7} k(Lseq_{p,s,k})^{k}$$

p,s = 1 to ∞

(9)

Illogical sequence (IS): $IS_{p, s} = ILseq_s, ILseq_s, ..., ILseq_{\infty}$ (10)

The separator outputs which are illogical in their category are fed to the discriminator (D2) where the discriminator output defined by

$$D2_{p,s} = ILseq_s^{m}$$
where $p, s, m = 1$ to ∞
(11)
 $m = Number of times nucleotide base is repeated$

The comparator unit compares the identification numbers of all logical sequences of mutated and non-mutated DNA inputs from Discriminator (D1) and illogical sequences of mutated and non-mutated sequences from (D2) to identify the location of mutation in the given sample.

DNA SAMPLE: *HUMAN-1* [BASE PAIR =32, SEQUENCE =25] AATGTGTTGTGTGACCCCTCAAAATCTCTCAAATGTGTTTTTACAC TCCGTTGGTAATATGGAATGTGTTAAAGTTGCTACCCGGGGTTTT TTAATGTGTCTCT

TABLE 4 DISCRIMINATOR (D1) OUTPUTS FOR NON-MUTATED LOGICAL SEQUENCE OF *HUMAN-1*

Sequ	Discr	Logical Sequence(LS _{p,s,k})	Identific	Uniq





Logi cal Sequ ence	Hu man (p)	ence (s)	imin- ator(D1) Input S (l _{p,s,k})	p,s,k	Lseq _{p,s,k} (k=2)	p,s,k	Lseq _{p,s,k} (k=4)	Lseq ^{p,s,k} (k=5)	Lseq p,s,k (k=6)	Lseq ^{p,s,k} (k=7)	ation Number (D1 _{p,s})	ue Identi ficat -ion numb er (UIN _p)
LS1	1	1	L 1, 1, k	0.1	0.1	0.2	0.3	0.2	0.3	0.2	0.18246 4	
LS2	1	2	L <u>1, 2,</u> k	0.2	0.3	0.2	0.3	0.2	0.3	0.1	0.44237 5	-
LS3	1	3	L 1, 3, k	0.4	0.2	0.4	0.1	0.1	0.1	0.1	0.67245 7	-
LS4	1	4	L 1, 4, k	0.2	0.4	0.2	0.4	0.2	0.4	0.1	0.67113 7	
LS5	1	5	L <u>1, 5,</u> k	0.1	0.1	0.2	0.3	0.2	0.3	0.2	0.18246 4	0.182
LS6	1	6	L 1, 6, k	0.2	0.1	0.4	0.1	0.4	0.2	0.4	0.47545 3	464 (REP
LS7	1	7	L 1, 7, k	0.4	0.3	0.2	0.2	0.3	0.3	0.2	0.62701 4	EAT ED
LS8	1	8	L 1, 8,	0.1	0.1	0.2	0.1	0.2	0.3	0.3	0.15046 5	PAT TER
LS9	1	9	L 1, 9, k	0.1	0.1	0.2	0.3	0.2	0.3	0.2	0.18246 4	N)
LS1 0	1	10	L 1,10,k	0.2	6 0.1 c	0.1	0.1	0.3	0.2	0.2	0.23948 0	
LS1 1	1	11	L 1,11,k	0.3	0.4	0.2	0.1	0.4	0.4	0.4	0.73164 5	
LS1 2	1	12	L 1,12,k	0.3	0.2	0.2	0.2	0.2	0.2	0.2	0.41103 3	
LS1 3	1	13	L 1,13,k	0.1	0.1	0.2	0.3	0.2	0.3	0.2	0.18246 4	

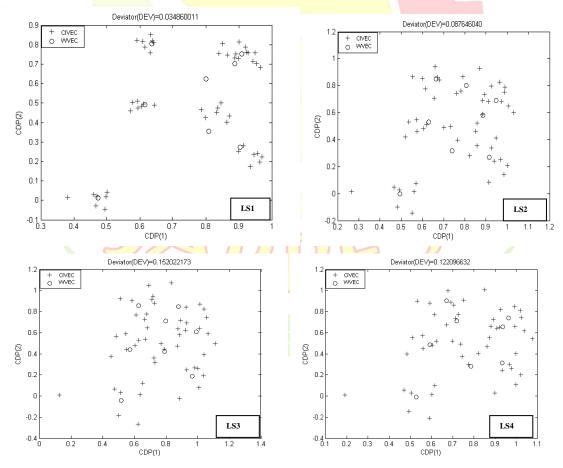


TABLE 5

PERFORMANCE OF PROPOSED SYSTEM FOR VARIOUS NUMBERS OF EPOCHS

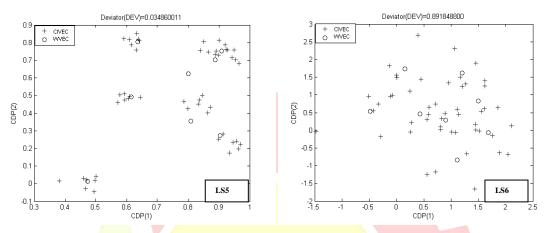
Vigila	Vigilance Parameter (p):0.5											
S. no	Learning Vector (Number of Epochs)	Number of Learning	Learning Time	Accuracy%								
		Inputs	(Seconds)									
1	25	25	62.49	100%								
2	13	13	34	100%								

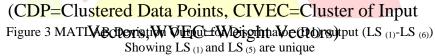
(No. of Epochs=25(For All Possible Combinations) and No. of Epochs=13)





International Journal of Advanced Research in Biology Ecology Science and Technology (IJARBEST) Vol. 2, Special Issue I, January 2016 in association with KARUR VELALAR COLLEGE OF ARTS AND SCIENCE FOR WOMEN, KARUR Conference on Emerging Trends and Functional Applications of Computer Technology - 12th January 2016





5. IDENTIFCIATION OF MUTATION IN THE SAMPLE

Mutation is a change of DNA sequence within a gene or chromosome of an organism resulting in the creation of a new character or trait not found in the parental type [22]. The mutation results when a change occurs in a chromosome, either through an alteration in the nucleotide sequence of the DNA coding for a gene or through a change in the physical arrangement[24] [25] of a chromosome.

There are many different types of mutations; a point mutation (base pair substitution) is a simple change in one base of the gene sequence. In this case, the entire meaning of the sentence has been altered with a one letter change. In neutral or silent mutation, another one letter point mutation has occurred. However, the meaning of the sentence has not been altered.





In a frame shift mutation, one or more bases are inserted or deleted into the sequence of the gene, the equivalent of adding or removing letters in a sentence, adding or removing one letter changes each subsequent word. This type of mutation can make the DNA meaningless and often results in shortened and functionless protein. Mutations that result in missing DNA are called deletions. These can be small, or longer deletions that affect a large number of genes on the chromosome. Deletions can also cause frame-shift mutations. Mutations that result in the addition of extra DNA are called insertions. Insertions can also cause frame-shift mutations, and generally result in a nonfunctional protein.

In an inversion mutation, an entire section of DNA is reversed. A small inversion may involve only a few bases within a gene, while longer inversions involve large regions of a chromosome containing several genes.

Before Mutation:						
LS1/RS LS2 IS1 IS1 IS1 LS3 LS4						
LS5/RS IS2 IS2						
AATGTGT TGTGTGA C C C CTCAAAA						
TCTCTCA AATGTGT T T T						
IS2 LS6 LS7 LS8 LS9/RS LS10						
LS11 IS3 IS3						
T TACACTC CGTTGGT AATATGG AATGTGT TAAAGTT						
GCTACCC G G						
IS3 LS12 LS13/RS LS14						
G GTTTTTT AATGTGT CTCTXXX						
Case 1:-						
After Mutation in Logical sequence: [Point mutation]						
34						
34						

5.1 Various Types of Mutation identification in Human-1 sample





International Journal of Advanced Research in Biology Ecology Science and Technology (IJARBEST) Vol. 2, Special Issue I, January 2016 in association with						
KARUR VELALAR COLLEGE OF ARTS AND SCIENCE FOR WOMEN, KARUR Conference on Emerging Trends and Functional Applications of Computer Technology - 12 th January 2016						
LS1/RS	LS2	IS1	IS1	IS1	LS3	LS4
LS5/RS IS2	IS2					
AATGTGT TG	IGTGA	С	С	С	CTCA C AA	
TCTCTCA AATGTGT T T						
IS2 LS6	LS7		L	. S8	LS9/RS	LS10
LS11 IS3	IS3					
T TACACTC	CGTTGC	GT A	АТА	TGG	AATGTGT	TAAAGTT
GCTACCC G	G					
IS3 LS <mark>12</mark>	LS13/RS		LS14			
G GTTTT	AATGTG7	Г СТО	CTX	XX		

In case 1 the point mutation occurred in logical sequence $(LS_{1,3})$ by the mutant C can be identified with the change in identification number of $LS_{1,3}$ where identification number of illogical sequence remains unaltered.

Logical Sequence (LS)	Repeated Sequence (RS)	Identification Number	DES
$LS_{1,1}$	RS	0.182464	REE
LS _{1,2}		0.442375	
$LS_{1,3}$	R	0.672457	
LS _{1,4}		0.671137	its Best III
LS _{1,5}	RS	0.182464	
LS _{1,6}		0.475453	
LS _{1,7}		0.627014	
LS _{1,8}		0.150465	
LS _{1,9}	RS	0.182464	
LS _{1,10}		0.239480	
LS _{1,11}		0.731645	
LS _{1,12}		0.411033	



$\mathbf{LS}_{1,13}$ NS 0.102404 Define with an observe of the second se	$LS_{1,13}$	RS	0.182464	Before Mutation
--	-------------	----	----------	------------------------

After [Point Mutation] in Logical Sequence

<u>Before Mutation</u> <u>After Mutation in logical Seque</u>	Logical Repeated Sequence Sequence (LS) (RS) Identification Number	
(Not Altered)	LS_{1,1} RS 0.182464	
Illogical Linetic Line	LSHogical 0.442375	<
Sequence Identification	LStauence Identification 23607	
(IS) Number	LS ₁ ,(IS) Number 0.671137	
$IS_{1,1}$ 0.064000	LS _{1,IS} _{1,1} RS _{0.06400} 0.182464	
1000000000000000000000000000000000000	LS _{1,JS_{1,2} 0.008000.475453}	
$1S_{1,3}$ 0.027000	LS _{1,JS1,3} 0.027000.627014	
	LS _{1,8} 0.150465	
	LS _{1,9} RS 0.182464	
Degulte Change in nolymon	LS _{1,10} 0.239480	
[Result: Change in polypep	10e 0.731645	
sequence might change the shape function of the protein, depending	01 - 0.411033	
where in the sequence occurs	LS_{1,13} RS 0.182464	

where in the sequence occurs]

Case 2:- <u>After M</u>	Iutation :	in Logical s		e: [Fr	ame shif) t mutation	n-Insertion]
LS1/RS	5	LS2 sea	IS1	IS1	IS1 _{est}	LS3	LS4
LS5/RS	IS2	IS2					
AATGT	GT TO	GTGTGA		С	С	С	CTCAAAA
тстсто	CA AA	TGTGT '	Т Т				
IS2	LS6	LS7		LS8		LS9/RS	LS10
LS11	IS3	LS12					



T TACACTC CGTTGGT AATATGG AATGTGT TAAAGTT GCTACCC G CGGGTTT IS4 1S4 LS13 LS14 C

T T TAATGTG TCTCTXX

In case 2 the frame shift mutation (insertion) occurred in one of the $IS_{1,3}$ by the mutant C which alters both the logical sequence ($LS_{1,12}$) and illogical sequence ($IS_{1,3}$) that can be identified by the change in identification number of both logical sequence ($LS_{1,12}$) and illogical sequence ($IS_{1,3}$) after mutation.

Before Mutation

After Mutation in

Logical Sequence (LS)	Repeated Sequence (RS)	Identificatio Number	n Logical Sequence (LS)	Repeated Sequence (RS)	Identification Number
$LS_{1,1}$	RS	0.182464	LS _{1,1}	RS	0.182464
$LS_{1,2}$	115	0.442375	LS _{1,2}		0.442375
$LS_{1,3}$		0.672457	LS _{1,3}	nG V	0.672457
LS _{1,4}	11	< 0.671137	LS1,4		0.671137
LS _{1,5}	RS	0.182464	LS1,5	RS	0.182464 Mutat
$LS_{1,6}$		0.475453	LS1,6		0.475453 occurr after LSt.
LS _{1,7}	R	0.627014	LS _{1,7}		0.627014
LS _{1,8}		0.150465	LS1,8	SU	0.150465
LS _{1,9}	RS	0.182464	LS _{1,9}	RS	0.182464
LS _{1,10}		0.239480	LS _{1,10}		0.239480 ♦
LS _{1,11}		0.731645	LS _{1,11}		0.731645
LS _{1,12}		0.411033	LS _{1,12}		
LS _{1,13}	RS	0.182464	LS _{1,13}		0.243464

<u>Illogical Sequence</u>

Before Mutation Illogical Sequence After Mutation in



Illogical Sequence (IS)	Identification Number		Illogical Sequence (IS)	Identification Number	Mutation occurred in IS _{1,3}
IS _{1,1}	0.064000		IS _{1,1}	0.064000	
IS _{1,2}	0.008000		IS 1,2	0.008000	
IS 1,3	0.027000	[Result:	IS 1,3		
	1	-	IS ₁₄	0.040000	

Change in polypeptide sequence might

change the shape or function of the protein, depending on where in the sequence occurs.]

Case 3:-

Silent)]

Silene)]				
LS1/RS	LS2	IS1 IS1 IS1	LS3	LS4
LS5/RS	IS2 IS2 IS2			
AATGTGT	TGTGTGA	C C C	CTC AAAA	TCTCTCA
AATGTGT	ТТТ			
LS6	LS7	LS8	LS9/RS	LS10
LS11	IS3 IS3	mm	2GV	
TACACTC	CGTTGGT	AATATGG	AATGTGT	TAAAGTT
GCTACCC	G G			
IS3 IS	3 LS12 Ca	LS13/RS	LS14	
G G	GTTTTTT	AATGTGT C	ГСТХХХ	G

Logical	Repeated	Identification	
Sequence	Sequence	Number	
(LS)	(RS)	Inumber	
LS _{1,1}	RS	0.182464	



Befo	re Mutatio	on	$LS_{1,2}$		0.442375
	After Mutation				0.672457
	AI		LS _{1,4}		0.672577
<u>in Illogical Sequence (</u> Not			LS _{1,5}	RS	0.182464
Altered)			LS _{1,6}		0.475453
1 XII.	icu)		LS _{1,7}	0.627014	
Logical	Repeated		LS _{1,8}		0.151905
Sequence	Sequence	Identification	LS _{1,9}	RS	0.182464
(LS)	(RS)	Number –	LS _{1,10}		0.236024
LS _{1,1}	RS	0.182464	LS _{1,11}		0.731645
LS _{1,2}		0.442375	LS _{1,12}		0.412474
LS _{1,3}		0.672457	LS _{1,13}	RS	0.182464
LS _{1,4}		0.671137			
LS _{1,5}	RS	0.182464	[Result: N	No change	e in polypeptide

[**Result:** No change in polypeptide sequence, possible consequence for the organism =none]

In case 3 the point mutation is occurred in same $IS_{1,3}$ as case 2 but with mutant G that only alters the illogical sequence ($IS_{1,3}$) and not any of the logical sequences that can be identified only using

the change in identification number of illogical sequence ($IS_{1,3}$).

0.475453

0.627014

0.150465

0.182464

0.239480

0.731645

0.411033

0.182464

Before Mutation

RS

RS

Illogical Sequence

 $LS_{1.6}$

LS_{1,7}

 $LS_{1.8}$

LS_{1,9}

LS_{1,10}

LS_{1,11}

LS_{1,12}

LS_{1,13}

After Mutation in

Mutation occurred in IS_{1,3}

	Rese		Best III		_
Illogical Sequence (IS)	Identification Number		Illogical Sequence (IS)	Identification Number	$\left(\right)$
IS _{1,1}	0.064000		IS _{1,1}	0.064000	_
IS _{1,2}	0.008000		IS _{1,2}	0.008000	-
IS _{1,3}	0.027000	Case 4:-	IS _{1,3} ⁻		-

<u>After Mutation in Logical sequence</u>: [Frame shift mutation-Deletion]



International Journal of Advanced Research in Biology Ecology Science and Technology (IJARBEST) Vol. 2, Special Issue I, January 2016 in association with							
				NCE FOR WOMEN, Computer Technolo	KARUR pgy - 12 th January 2016		
LS1/RS	LS2	IS1 IS	1 IS1	LS3	LS4		
LS5/RS IS2	2 IS2						
AATGTGT T	GTGTGA	С	С	С	CTCAAAA		
TCTCTCA A	ATGTGT	ТТ					
IS2 LS6	LS7		LS8	LS9/R	S LS10		
LS11 IS3 IS3							
T TACACTO	C CGTTGG	GT AA	FATGG	AATGTGT	TAAAGTT		
GCTACCC G	G						
IS3 LS <mark>12</mark>	LS13/RS		Т	LS14			
G GTŢT <mark>T</mark> TA	ATGTGT	С [TCTXXX			

In case 4 the frame mutation [deletion] occurred in logical sequence $(LS_{1, 12})$ by the removal of mutant T and can be identified with the change in identification number of logical sequence $(LS_{1, 12})$ with no alteration in any of the illogical sequence.

<u>Before N</u>	<u>Iutation</u>	Sequence	After Mutation in Logical
Logical Sequence (LS)	Repeated Sequence (RS)	Identification Number	Dest III
LS _{1,1}	RS	0.182464	
LS _{1,2}		0.442375	
LS _{1,3}		0.672457	
LS _{1,4}		0.671137	
LS 1,5	RS	0.182464	
LS1,6		0.475453	
LS _{1,7}		0.627014	
LS _{1,8}		0.150465	
LS1,9	RS	0.182464	



LS _{1,10}		0.239480	Logical	Repeated	T.1
LS _{1,11}		0.731645	Sequence	Sequence	Identification
LS _{1,12}		0.411033	$\overline{(LS)}$	(RS)	Number
LS _{1,13}	RS	0.182464	LS _{1,1}	RS	0.182464
			LS _{1,2}		0.442375
Bofo	ro Mutotio		LS1,3		0.672457
<u>Before Mutation</u>			LS _{1,4}		0.67113 Mutation
<u>After Mutation in</u>			LS _{1,5}	RS	0.18246 occurred after LS1,
<u>Illogical Sequence</u> (Not Altered)			LS _{1,6}		0.475453
			LS _{1,7}		0.627014
Illogical	Identificat	ion	LS _{1,8} Ille		ntification65
Sequence	Number	N	LS _{1,9} Seq	uenges	Numb182464
(IS)	Indiliber		LS _{1,10}	(IS)	0.239480
IS _{1,1}	0.064000	0		. <mark>S_{1,1} (</mark>	0.0640991645
IS _{1,2}	0.008000] [Resu		S _{1,2} =C	1.0080200945=
IS 1,3	0.02700) lt:	LS _{1,13}	.S _{1,3} ()	0.0270991402

Change in polypeptide sequence

might change the shape or function of the protein, depending on where in the sequence occurs.]

In case 5 below the inversion mutation occurred in logical sequence $(LS_{1, 10})$ by replacing TAAAGTT with mutant TTGAAAT that can be identified with the change in identification number of logical sequence $(LS_{1, 10})$ alone with no alteration in any of the illogical sequence.

Case 5:-

After Mutation in Logical sequence: [Inversion mutation]						
LS1/RS	LS2	IS1	IS1	IS1	LS3	LS4
LS5/RS	IS2 IS2					
AATGTGT	TGTGTGA		С	С	С	CTCAAAA
TCTCTCA	AATGTGT	Т	Т			





152	L30	LS/	L38	L39/K3	b LSI	0
LS1	I IS3	IS3				
Т	TACACTC	CGTTGGT	AATATGG	AATGTGT	TTGAAA	Т
GCT IS3	TACCC G LS12					
LS14		LSTS/KS	ΤΤ	G A	A A	Т
G	GTTTTTT	AATGTGT	CTCTXXX			

<u>Before Mutation</u>

After Mutation in

Logical Sequence

Logical Sequence (LS)	Repeated Sequence (RS)	Identification Number	Logical Sequence (LS)	Repeated Sequence (RS)	Identification
$LS_{1,1}$	RS	0.182464	LS _{1,1}	RS	0.182464
$LS_{1,2}$	~//>	0.442375	LS _{1,2}	26	0.442375
LS _{1,3}	00	0.672457	LS1,3	202	0.6724 Mutation occurred in
LS _{1,4}		0.671137	LS1,4	20-	0.671 LS1, 10
LS _{1,5}	RS	0.182464	LS _{1,5}	RS	0.182464
LS _{1,6}	$R_{\rm c}$	0.475453	LS _{1,6}	et 111	0.475453
LS _{1,7}		0.627014	LS _{1,7}		0.627014
LS _{1,8}		0.150465	LS _{1,8}		0.150465
LS _{1,9}	RS	0.182464	LS _{1,9}	RS	0.182464
LS _{1,10}		0.239480	LS _{1,10}	<	θ .3615 46
LS _{1,11}		0.731645	LS _{1,11}		0.731645
LS _{1,12}		0.411033	LS _{1,12}		0.411033
LS _{1,13}	RS	0.182464	LS _{1,13}	RS	0.182464



[Result: Change in polypeptide sequence might change the shape or function of the protein, depending on where in the sequence occurs.]

6. CONCLUSION

As an attempt to automate the genetic finger printing for precise identification of individuals from their DNA sample, the Neural-fuzzy Pattern Recognition System implemented using the concept of fuzzy resonance theory mapping discussed in the above work classifies the sequences to identify a unique number from the given sample that actually includes nucleotide basis of adenine, guanine, cytosine, thymine and uracil which are represented by fuzzy values respectively. Any type of mutation for instance, gene mutations in the Japanese HNPCC (Hereditary Non Polyposis Colorectal Cancer) which triggers HNPCC tumor that could not be detected even by PCR-SSCP(Polymerase chain reaction-Single strand conformation polymorphism) can be easily detected by subjecting the sample to gene sequencing process and analyzed using the proposed system.

Further development can be extended by training patterns in DNA protein represented by suitable fuzzy equivalent to classify and predict the protein structure in the protein folding problem and also above technique can be used in the areas where feature extraction is to be done in genetic engineering with suitable modification.

7. REFERENCES

- 1. Richard O. Duda, Peter E.Hart, David G. Stork, "Pattern classification"-Second Edition", John Wiley and sons, 2006.
- 2. John Hertz, Anders Krogh, and Richard G. Palmer. "Introduction to the Theory of Neural Computation". Addison Wesley, Redwood City, A, 2008.
- 3. Stephen J. Hanson, Jack D. Cowan, and C. Lee Giles, "Advances in Neural Information Processing Systems", volume 5, Morgan Kaufmann San Mateo CA, 2009.



International Journal of Advanced Research in Biology Ecology Science and Technology (IJARBEST) Vol. 2, Special Issue I, January 2016 in association with

KARUR VELALAR COLLEGE OF ARTS AND SCIENCE FOR WOMEN, KARUR

- Conference on Emerging Trends and Functional Applications of Computer Technology 12th January 2016
- 4. "Advances in Neural Networks issn-2006", Third international symposium on neural networks, Springer Berlin Heidelberg, New York publications.
- 5. Robert Schalkoff, "Pattern Recognition: Statistical, Structural and Neural Approaches, 2007, John Wiley and sons.
- 6. Carpenter, G.A. and S. Grossberg, "A Massively Parallel Architecture for a self-organizing Neural Pattern Recognition Machine", Computer Vision, Graphics and Image Processing, 37, PP. 54-115.
- Carpenter, G.A. and S. Grossberg, and J.H. Reynolds (2010), "ARTMAP: Supervised Real Time Learning and Classification of Nonstationary Data by a Self- organizing Neural Network". Vol. 4, pp. 565-588.
- 8. Phipps Arabie, Lawrence J. Hubert, and Geert De Soete, editors, "Clustering and Classification". World Scientific, River Edge, NJ.
- 9. Stephen, Krawetz, David D.Womble, "Introduction to Bioinformatics A Theoretical and Practical Approach", Human Press Inc.,
- 10.David W.Mount, David W. Mount, "Bio informatics Sequence and Genome analysis"- Second Edition, Cold Spring Harbor Laboratory Press, New York.
- 11.Norah Rudin, Keith Inman, "An Introduction forensic DNA Analysis", 2002-CRC Press.
- 12.Donald R. Tveter. "The Pattern Recognition Basis of Artificial Intelligence". IEEE Press, New York, page 117, Computational Intelligence and Bio inspired Systems, 8th international work conference on artificial neural networks, iwann-2005proceedings.
- 13.Julie A. Ayala-Gross, "DNA Analysis: The best method for Human Identifications", National University, San Diego 2001.
- 14.Joe Nickell and John F.Fischar, "Crime Science Methods of Forensic Detection", 1999. University Press of Kentucky.
- 15.John O. Savino, Brent E Turvey, "Rape Investigation Hand book", 2005, Elsevier Inc.,
- 16.David E. Newton, "DNA Evidence and Forensic science"- 2008 facts on file, Inc. http://www.factsonfile.com.
- 17. Jorg T. Epplen Thomas Lubjuhn, Birkhauser, "DNA Profiling and DNA Finger Printing", Verlag Publication.



International Journal of Advanced Research in Biology Ecology Science and Technology (IJARBEST) Vol. 2, Special Issue I, January 2016 in association with

KARUR VELALAR COLLEGE OF ARTS AND SCIENCE FOR WOMEN, KARUR

Conference on Emerging Trends and Functional Applications of Computer Technology - 12th January 2016

- 18.Simon Eastaeal, Neil Mc Lead, Ken, Harwood, "DNA Profiling Principles, Pitfalls and Potential", Academic Publishers, Inc.,
- 19.Des Higgins, willie Taylor, "Bioinformatics Sequence, Structure and data banks", Oxford University Press, 2000.
- 20."Bioinformatics for geneticists", Michael R.Barnes, Second Edition, John Wiley & Sons Ltd.,
- 21.Andreas D. Buxevanis, "Bioinformatics-A practical Guide to the Analysis of genes and proteins", second edition, A John wiley & sons, Inc., Publication.
- 22.Charles L.Valon, "New developments in Mutation Research", Nova science publishers Inc New York, 2007.
- 23."Oxidative Damage to Nucleic Acids", Springer science press, New York.
- 24.Richard G. H. Cotton, Edward Edkins, Sue Forrest "Mutation detection", IRL Press at Oxford University Press.
- 25.Graham R. Taylor "Laboratory methods for the detection of mutations and polymorphisms in DNA", CRC Press, 2007 - Science.
- 26.S.N Sivanandam, "Introduction to neural networks and MATLAB-6.0", Tata McGraw-Hill publishing company, 2006

